

HPAE-PAD analysis of Carbohydrates

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Presentation Outline

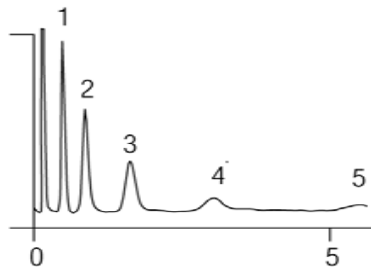
- ◆ Principles of High Performance Anion Exchange
- ◆ Column Technologies
- ◆ Principles of Pulsed Amperometric Detection
- ◆ Applications using HPAE-PAD

Carbohydrates - *Features of HPAE-PAD*

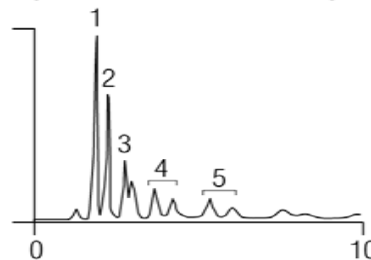
- ◆ **Simple, Direct Method**
 - No derivatization is necessary
- ◆ **High Sensitivity**
 - comparable to derivatization with fluorescence detection (fmol to low pmol).
 - Allows samples to be diluted 100 to 1000-fold, so simplifies sample prep
- ◆ **Minimal Sample Preparation is necessary**
- ◆ **Unique column technology**
- ◆ **Official Methods (using CarboPac PA1)**

Resolving Power of HPAE-PAD

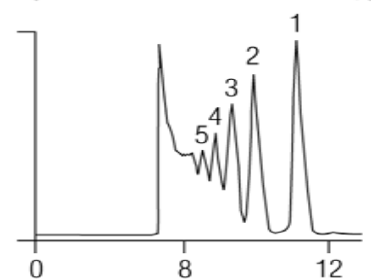
Oligo- and Polysaccharides Derived from Hydrolyzed Glucose Syrup



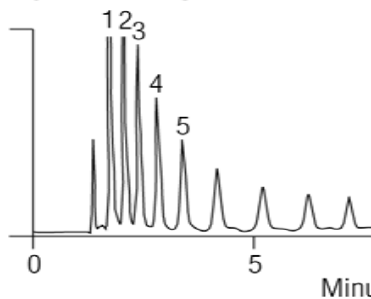
Column: Amino-Propyl
Eluent: 75% acetonitrile/
Water
Flow Rate: 2.0 mL/min
Detector: Refractive index
Temp.: 25 °C
Peaks: 1. DP-1
2. DP-2
3. DP-3
4. DP-4
5. DP-5



Column: Reversed-phase
Eluent: Water
Flow Rate: 0.5 mL/min
Detection: Refractive index
Temp.: Ambient



Column: Cation-exchange,
Calcium form
Eluent: Water
Flow Rate: 0.6 mL/min
Detector: Refractive index
Temp.: 80 °C



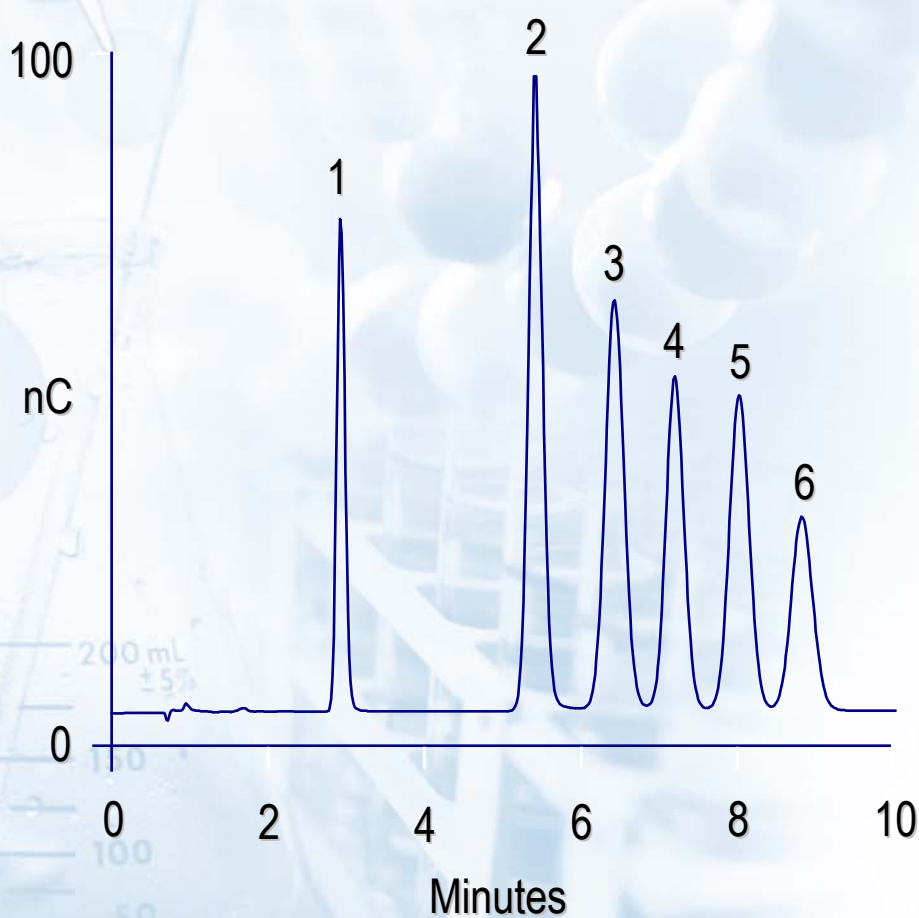
Column: CarboPac PA1
Eluent: Sodium hydroxide/
Sodium acetate
gradient
Flow Rate: 1.0 mL/min
Detector: Pulsed amperometry
Temp.: Ambient

Minutes

8576-8579

Comparison of chromatographic techniques used for the analysis of a hydrolyzed glucose syrup illustrates the remarkable resolving power of the HPAE-PAD technique. Elution order is reversed with the CarboPac® PA1 column as compared with conventional metal loaded cation-exchange columns.

Rapid Monosaccharide Analysis with the CarboPac™ PA10



Column: CarboPac PA10 and guard

Eluent: 18 mM NaOH

Flow Rate: 1.5 mL/min

Detection: Pulsed amperometry,
gold electrode

Peaks:

1. Fucose	1 nmol
2. Galactosamine	1
3. Glucosamine	1
4. Galactose	1
5. Glucose	1
6. Mannose	1

Detection of Food Sugars and Sugar Alcohols

Column: CarboPac™ PA10, PA10 guard,
Borate Trap®

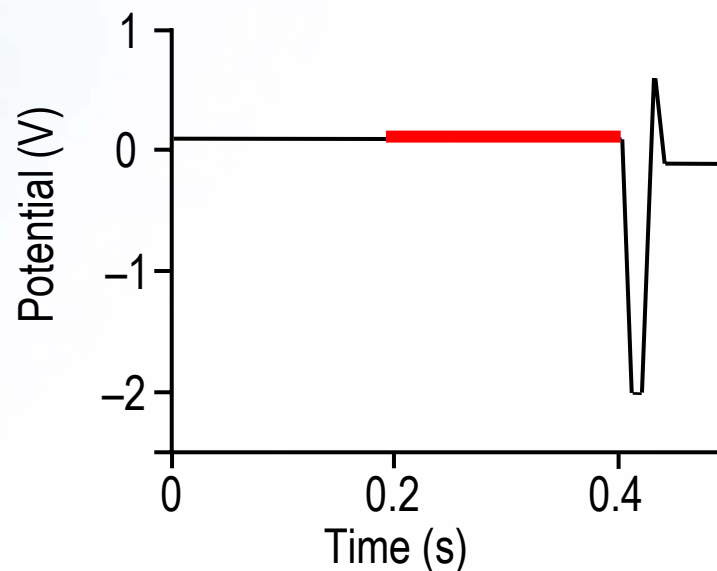
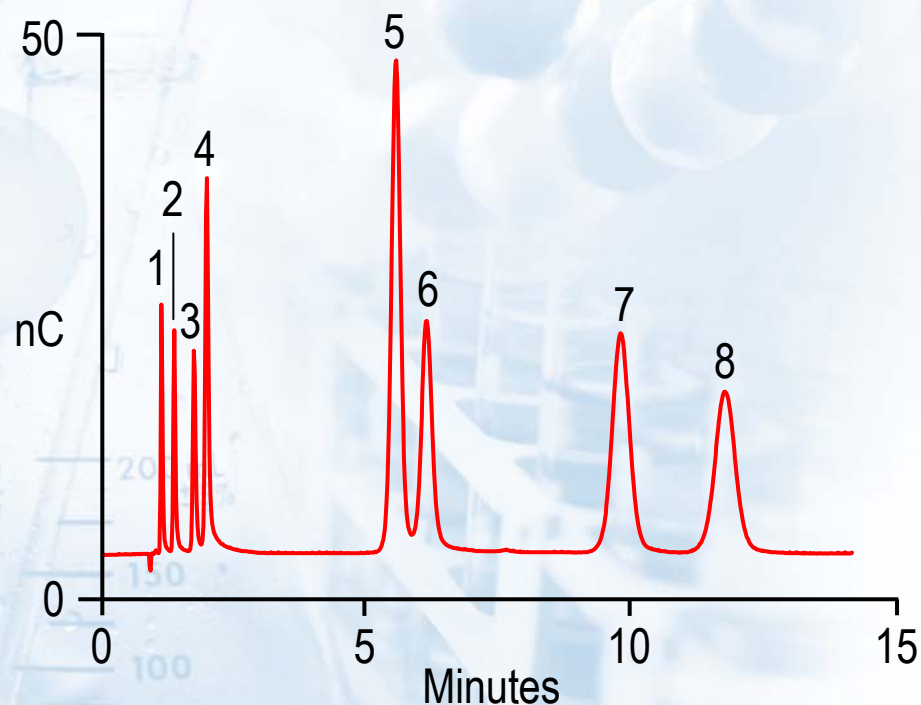
Eluent: 52 mM sodium hydroxide

Flow Rate: 1.5 mL/min

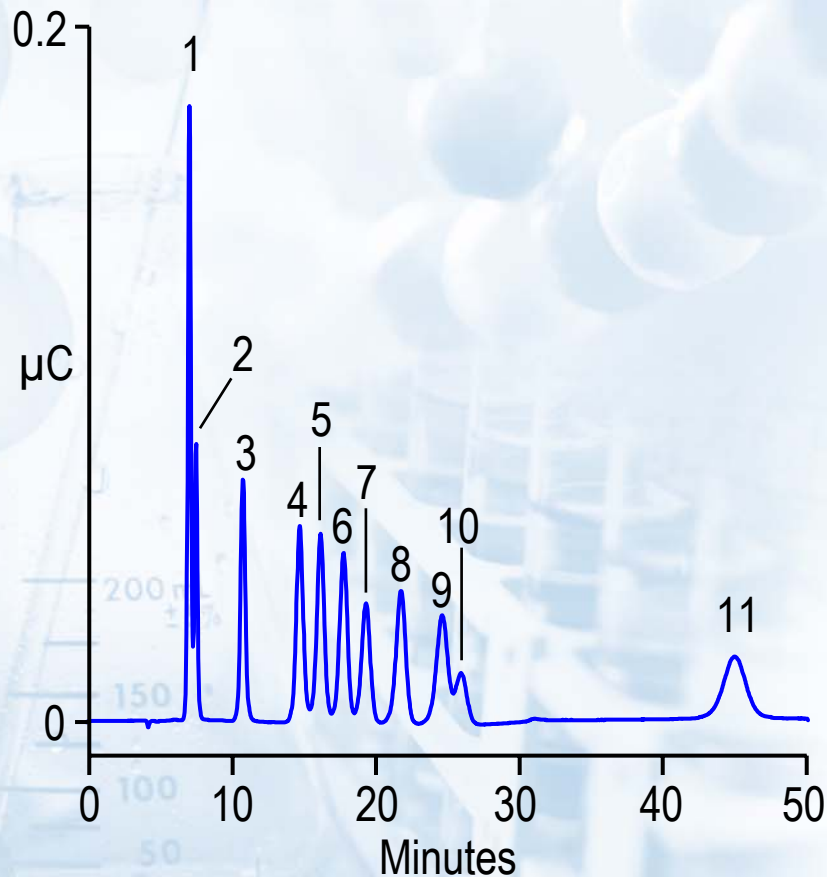
Inj. Volume: 5 μ L

Detection: Pulsed amperometry, gold electrode

Peaks:	1. Glycerol	4.0 mg/mL
	2. Xylitol	2.0
	3. Sorbitol	1.5
	4. Mannitol	3.0
	5. Glucose	5.0
	6. Fructose	8.0
	7. Sucrose	2.0
	8. Lactose	8.0



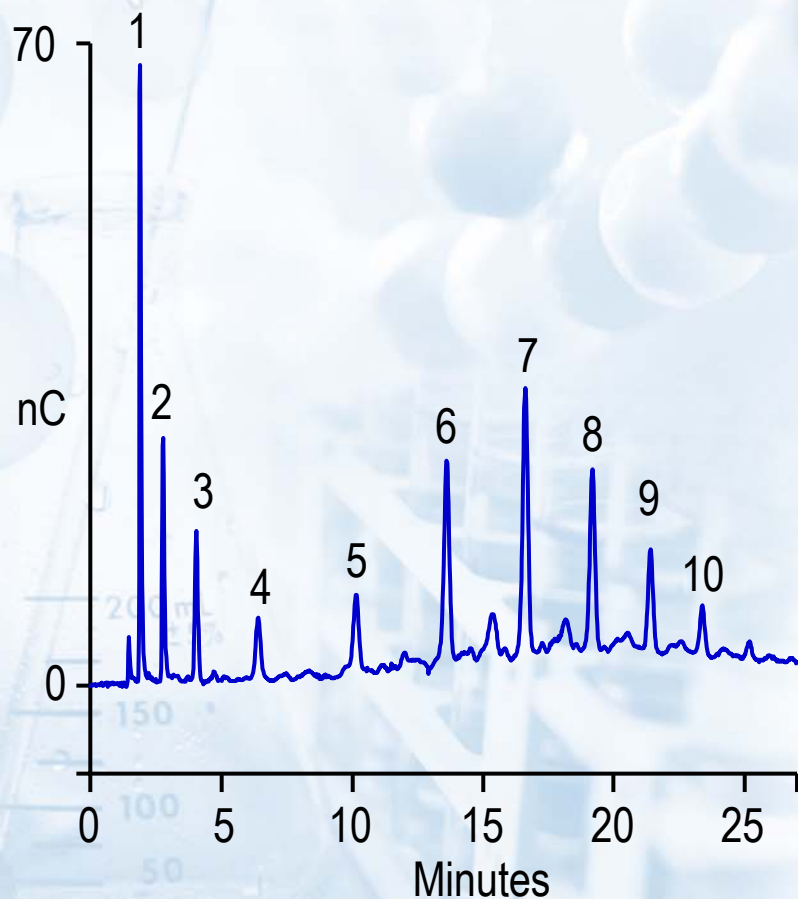
Separation of Reduced and Reducing Carbohydrates Commonly Found in Foods and Beverages



Column:	CarboPac™ MA1	
Eluent:	480 mM sodium hydroxide	
Flow Rate:	0.4 mL/min	
Detector:	PED (gold)	
Peaks:	1. Inositol	18 mg/L
	2. Glycerol	9
	3. Arabitol	15
	4. Sorbitol	18
	5. Dulcitol	18
	6. Mannitol	18
	7. Mannose	18
	8. Glucose	18
	9. Galactose	18
	10. Fructose	18
	11. Sucrose	34

Separation of Maltose Oligomers

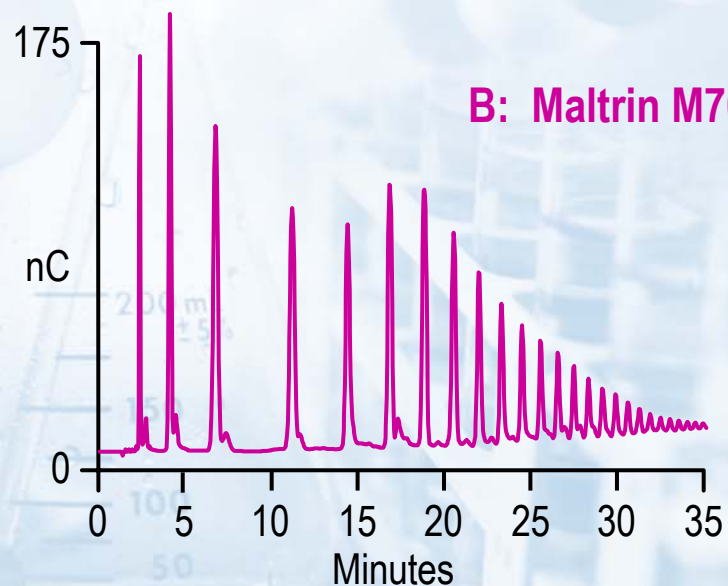
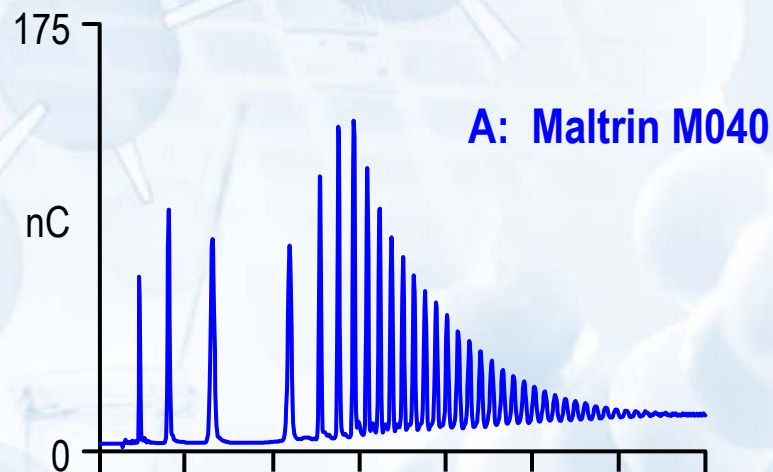
Column: CarboPac™ PA-100 and guard
Eluent: NaOH/sodium acetate gradient
Flow Rate: 1.0 mL/min
Inj. Volume: 10 μ L
Detection: Pulsed amperometry, Au electrode



Peaks:

1. Glucose
2. Maltose
3. Maltotriose
4. Maltotetraose
5. Maltopentaose
6. Maltohexaose
7. Maltoheptaose
8. Maltooctaose
9. Maltononaose
10. Maltodecaose

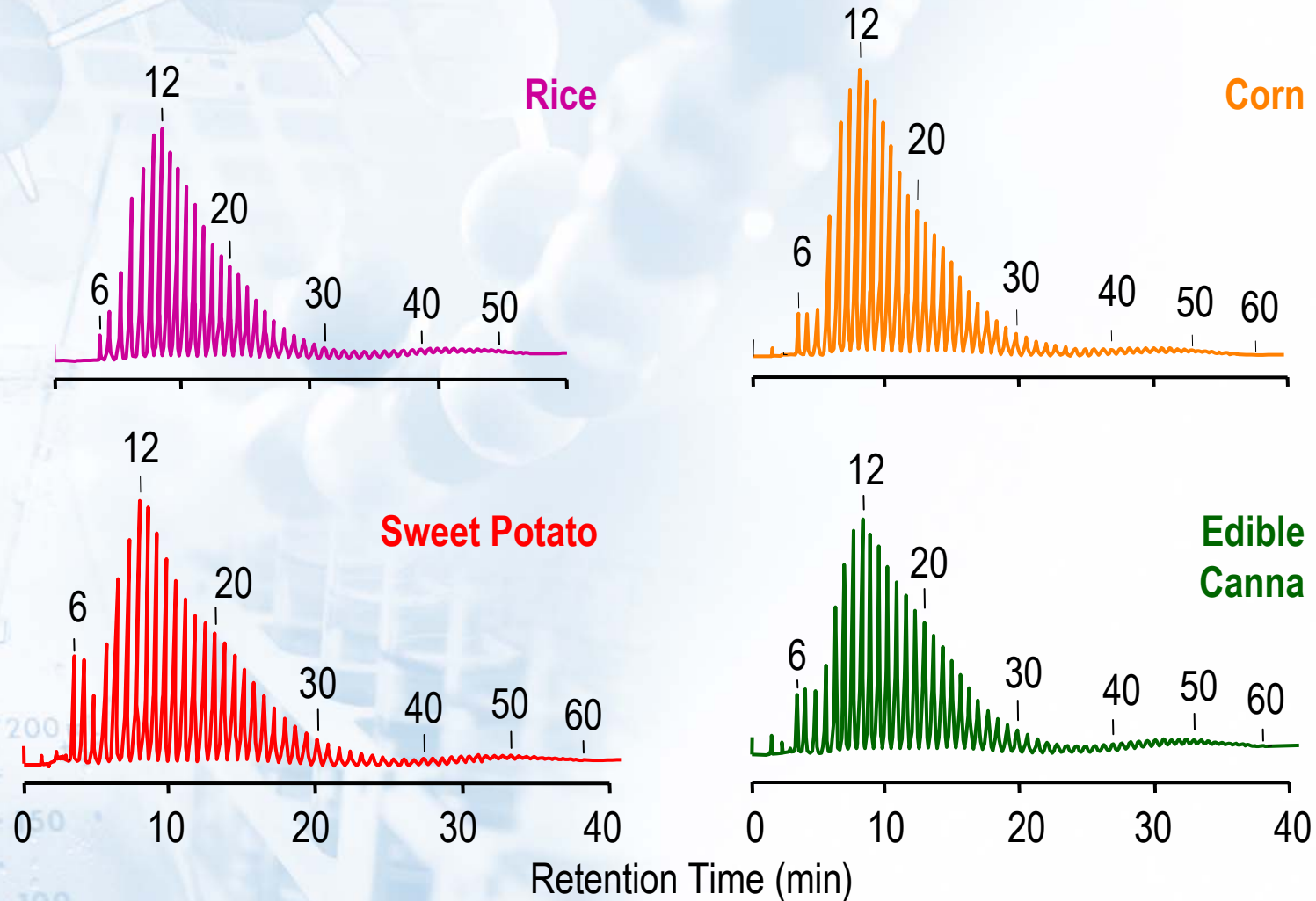
Maltodextrins—Maltrin[®] M040 and M700



Column: CarboPac[™] PA10
Eluent: Sodium hydroxide/
sodium acetate gradient
Flow Rate: 1 mL/min
Inj. Volume: 25 μ L
Detection: Pulsed amperometry,
Au electrode

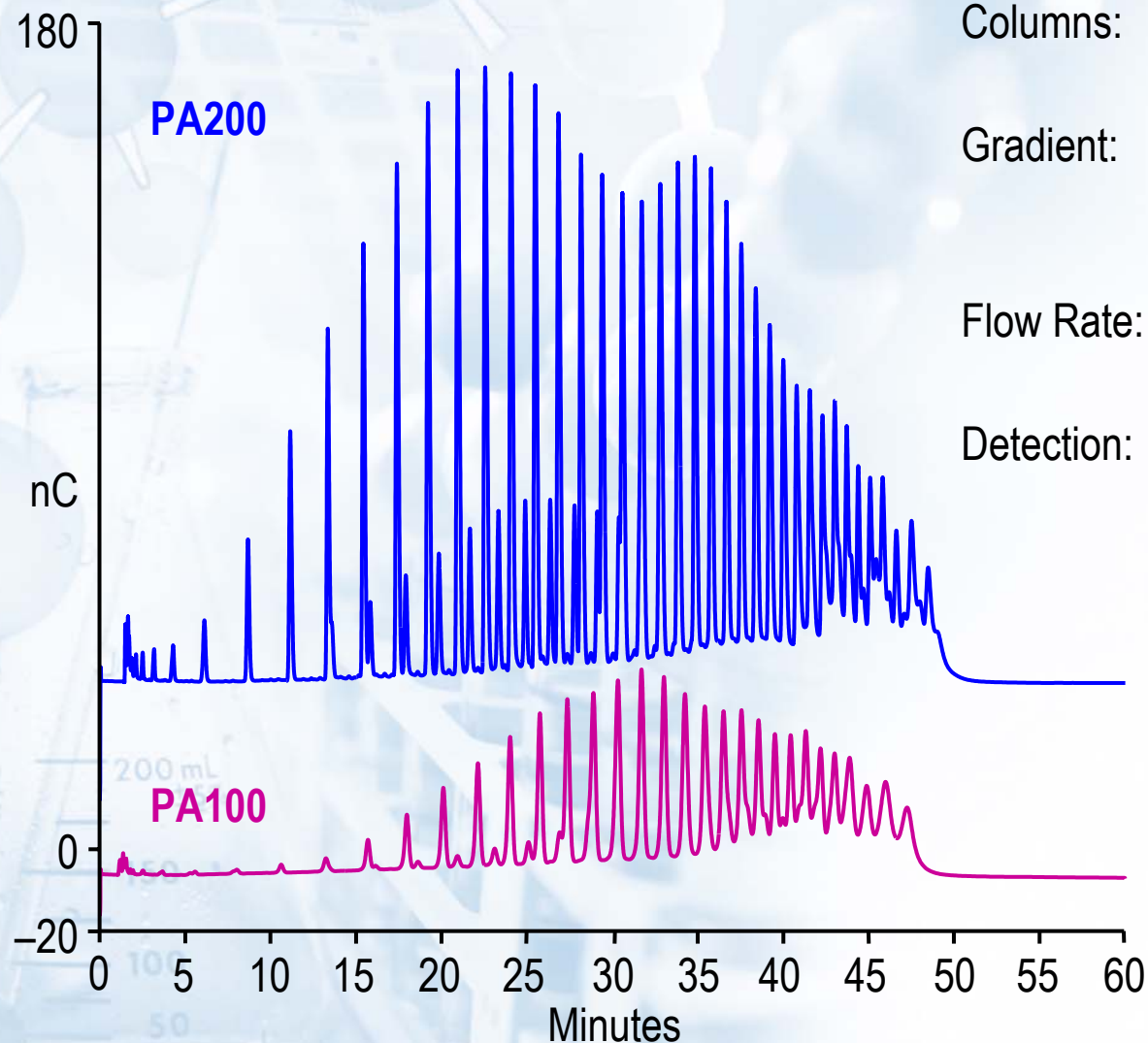
Maltrin is a trademark of Grain Processing Corp.

Distribution of Chain Length of Amylopectins by HPAE-PAD



From: Koizumi, K., Fukuda, M., Hizukuri, S.. *J. Chromatography*. **1991**, 585, 233–238

Improved Chain-Length Resolution of Inulin Polymers



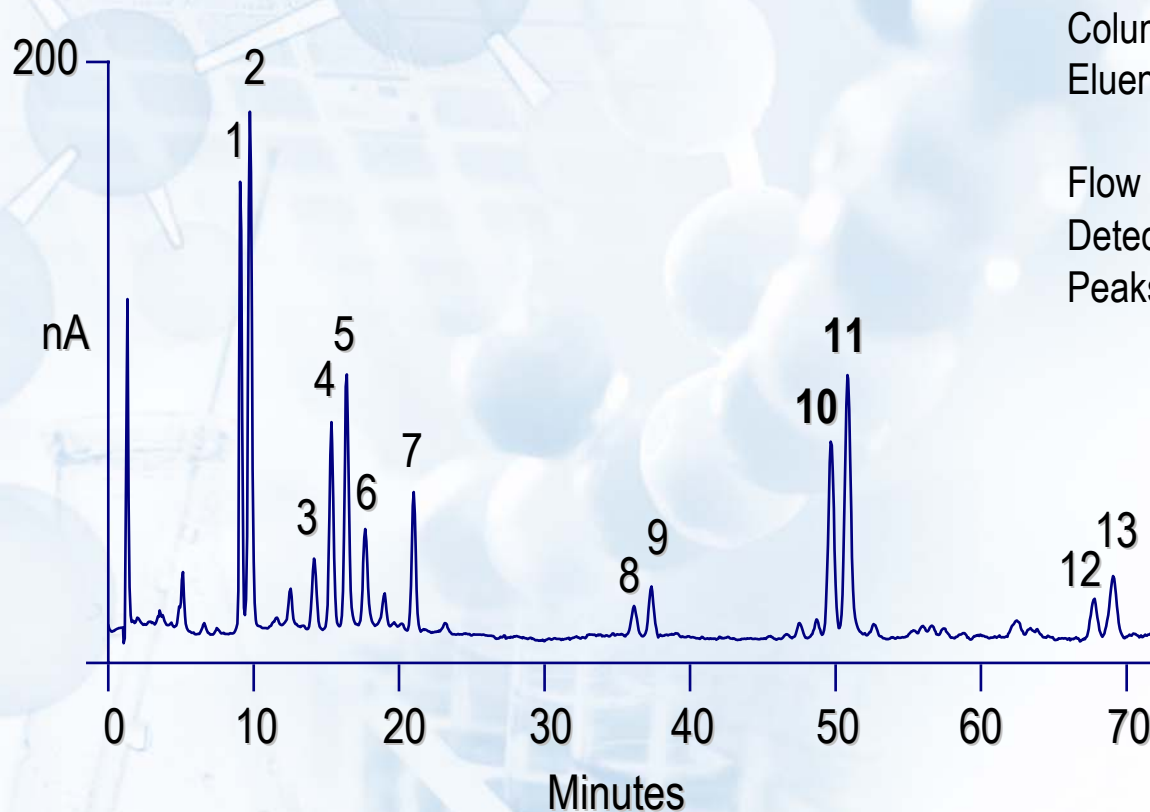
Columns: CarboPac™ PA200 (3 x 250 mm)
CarboPac PA100 (4 x 250 mm)

Gradient: 120- to 320-mM NaOAc
in 100 mM NaOH
over 40 min

Flow Rate: PA200: 0.5 mL/min
PA100: 1.0 mL/min

Detection: Pulsed amperometry,
quadruple waveform,
gold electrode

Separation of N-Linked Oligosaccharides



Column:

CarboPac™ PA-100

Eluent:

0 to 250 mM NaOAc over
110 min in 100 mM NaOH

Flow Rate:

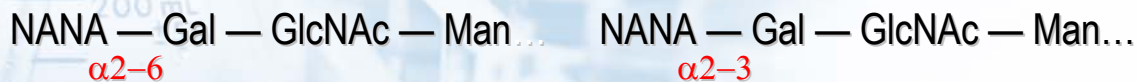
1 mL/min

Detector:

PAD (Gold)

Peaks:

1. Fucosylated Man₃GlcNAc₂
2. Man₃GlcNAc₂
3. Asialo, agalacto bi, core fuc
4. Asialo, agalacto bi
5. Asialo bi, core fuc
6. Asialo bi
7. Man₉GlcNAc₂
- 8, 9. Disialylated tri (reduced)
- 10, 11. Trisialylated tri (reduced)
- 12, 13. Tetrasialylated tri (reduced)



$\alpha 2-6$

$\alpha 2-3$

Peak 10

Peak 11

Official Food Methods Using IC/PAD

Analysis	CarboPac™ Column	Official Method
Sugars in Molasses	CarboPac PA1	AOAC Method 996.04 ICUMSA Method Approval 1994
Carbohydrates in Soluble Coffee	CarboPac PA1	AOAC Method 995.13 ISO 11292 Approved
Fructans in Food and Food Products	CarboPac PA1	AOAC Method 997.08
Polydextrose	CarboPac PA1	AOAC Method 2000.11
Transgalacto-oligosaccharides	CarboPac PA1	AOAC Method 2001.02
Low-Level Glucose and Fructose in Raw and Refined Sugar	CarboPac PA1	AOAC Method 2000.17



Column Technology

High Performance Anion Exchange Chromatography with Pulsed Amperometric Detection (HPAE-PAD)

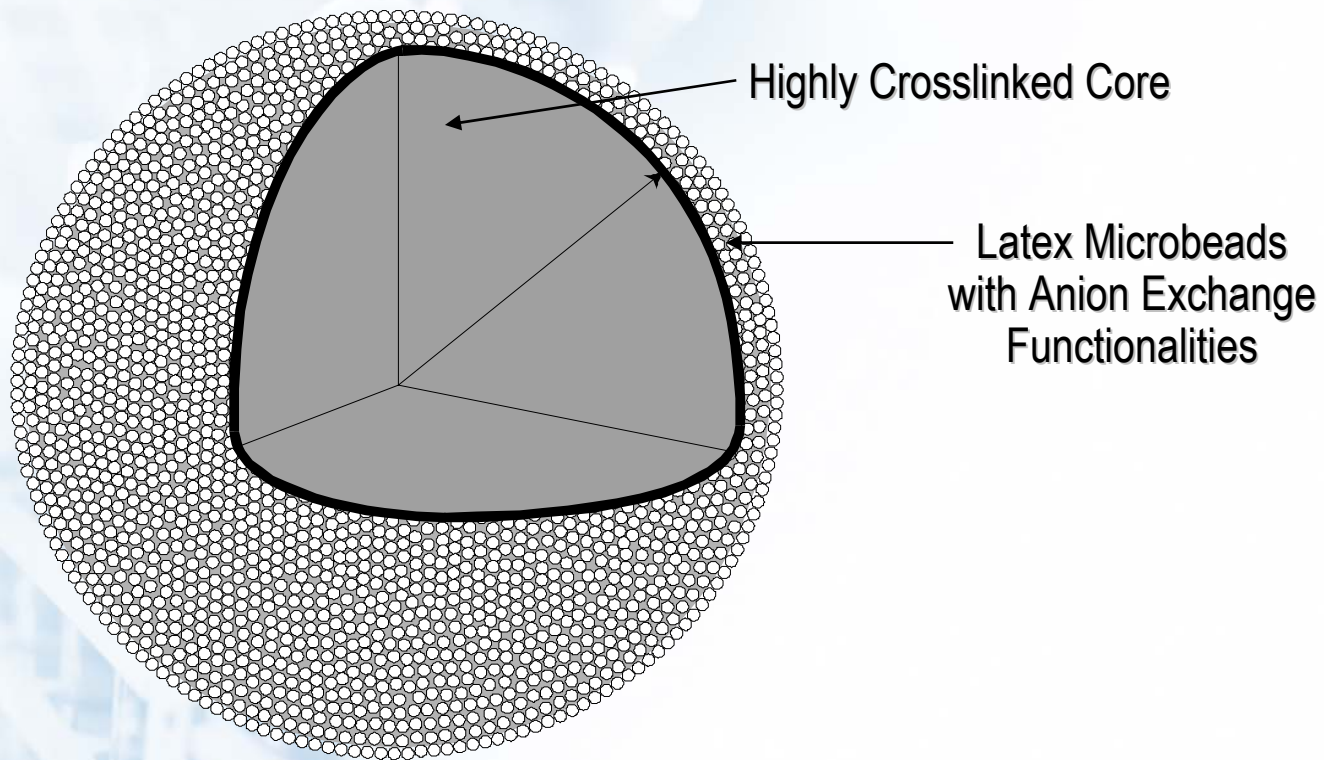
- ◆ Separates neutral and charged saccharides *without* derivatization
- ◆ Separates on the basis of:
 - Charge, size and composition
 - Branching, linkage isomerism

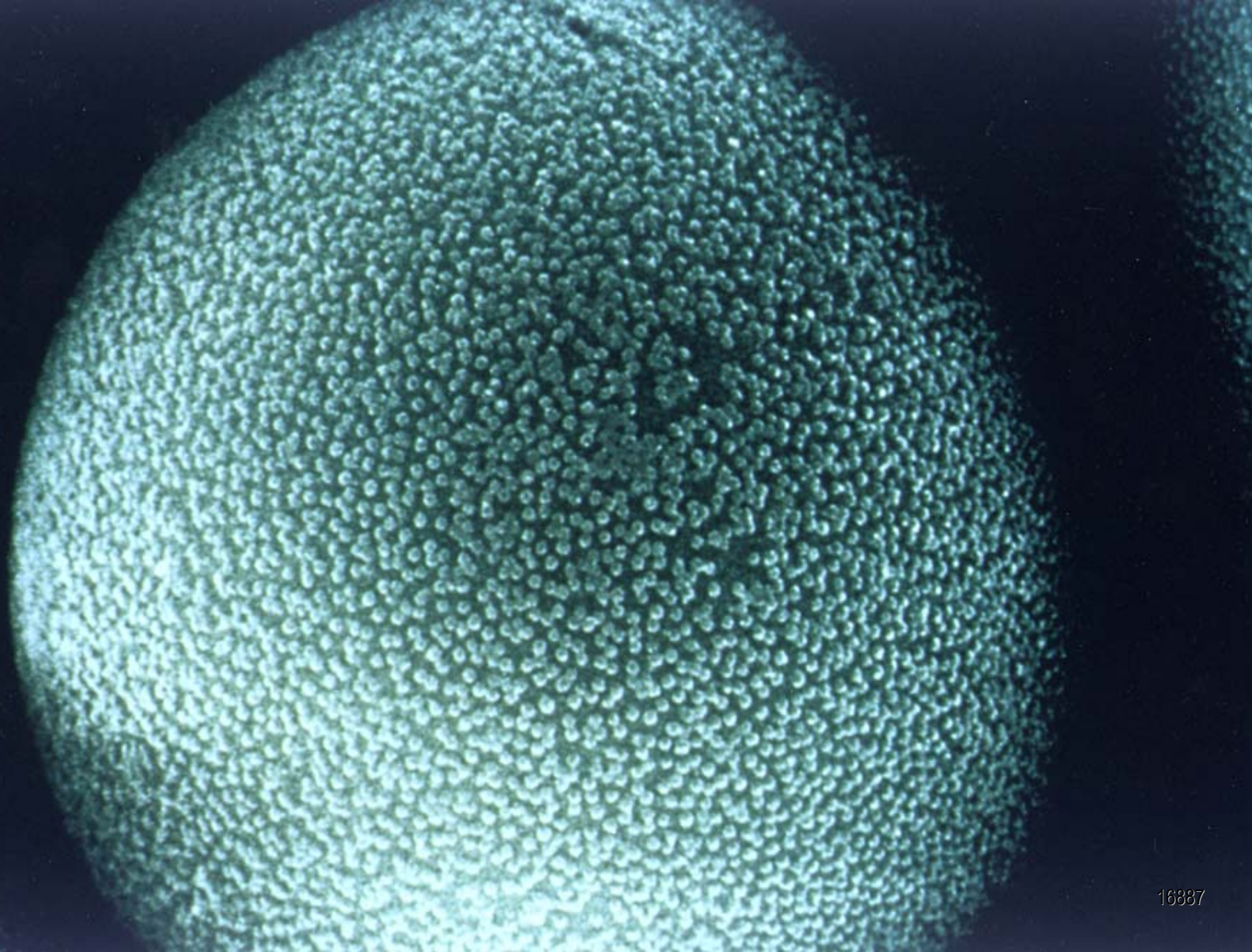
Dissociation Constants of Some Common Carbohydrates

(in Water at 25 °C)

Sugar	pK_a
Fructose	12.03
Mannose	12.08
Xylose	12.15
Glucose	12.28
Galactose	12.39
Dulcitol	13.43
Sorbitol	13.60
α -Methyl glucoside	13.71

Polymeric Nonporous (“Pellicular”) Packing







Electrochemical Detection

Pulsed Amperometry Detection

◆ Advantages of PAD for Carbohydrate Detection

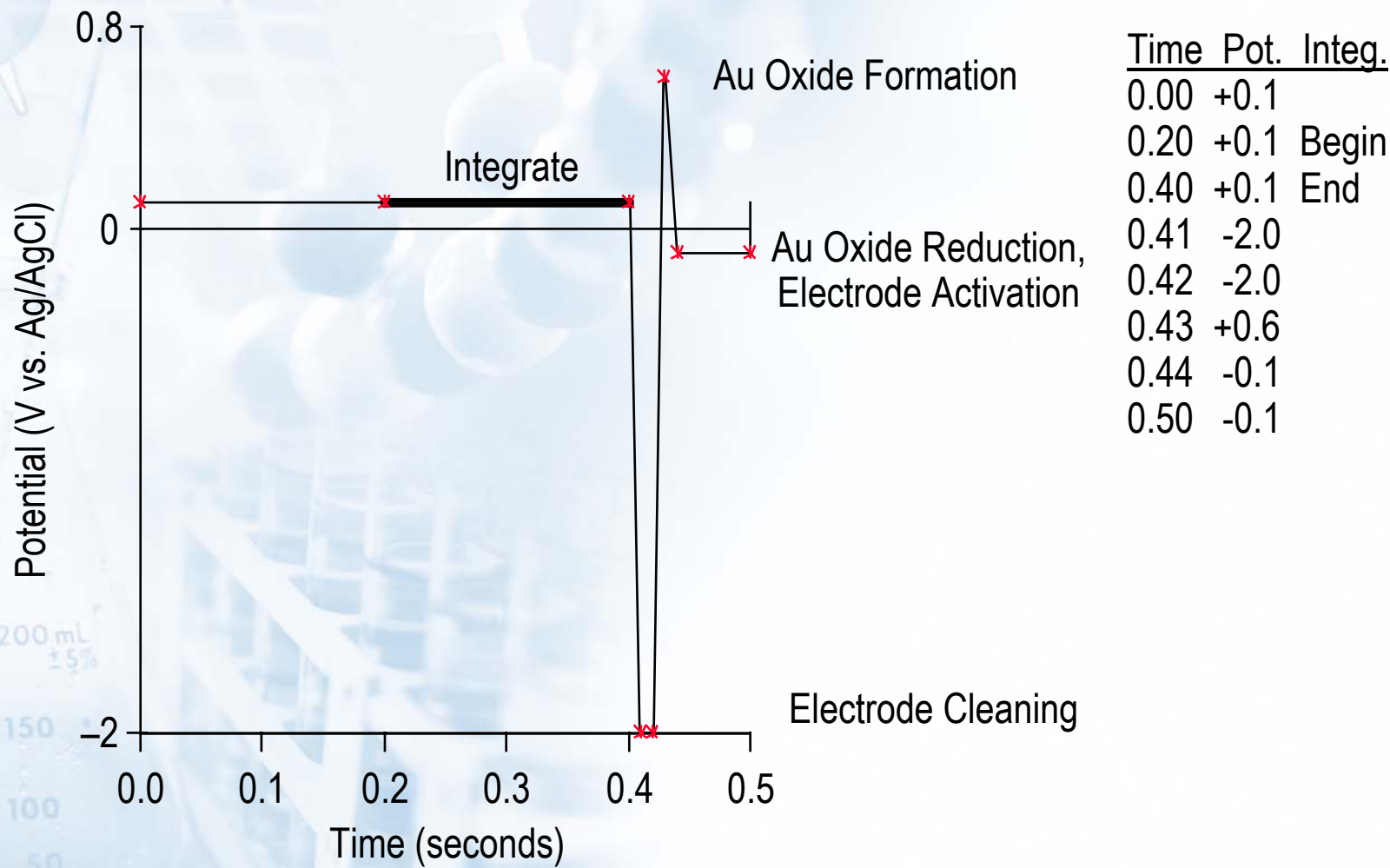
- No derivatization is necessary
- Low detection limits: low picomole to high femtomole
- Detection is linear over at least 4 orders of magnitude
- PAD is a non-destructive method and can thus be used preparatively
- Highly specific
- Using optimized pulse settings, PAD is highly specific for carbohydrates
- Versatile: isocratic or gradient elution, high pH or neutral pH (with post column base addition)
- Specific for electroactive species
- Both amino acids and carbohydrates can be detected in a single run

◆ Disadvantages of PAD for Carbohydrate Detection ***

- Each sugar gives a different signal, thus calibration curves must be run for quantitative results
- *** True for every quantitative analysis

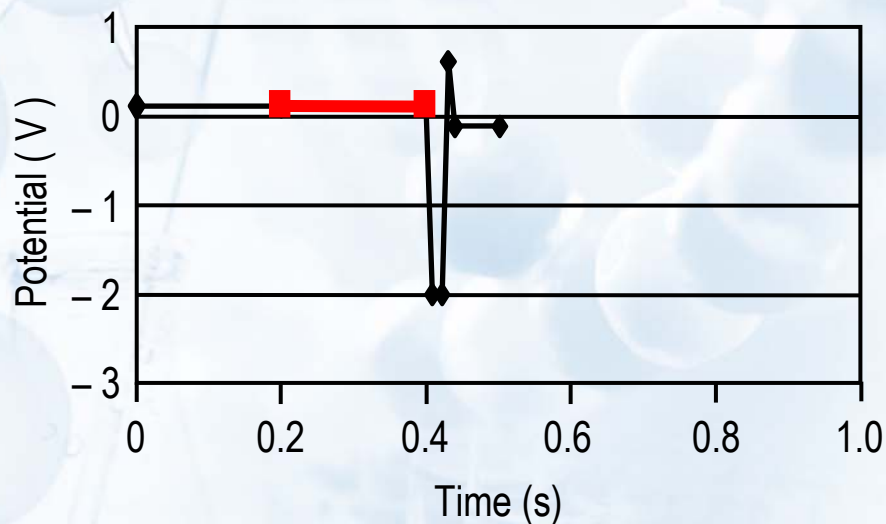
Principal of Pulsed Amperometric Detection (PAD)

Quadruple Potential Waveform for Carbohydrates

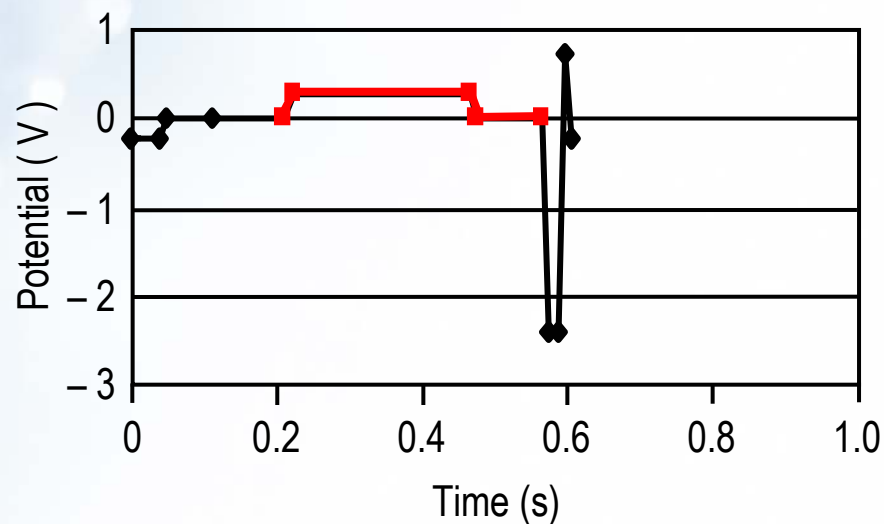


Pulsed Amperometric Detection (PAD)

Waveform and Integration Period



Carbohydrate Waveform



Amino Acid Waveform

— Integration Period

Sugars in Sugar Cane Molasses

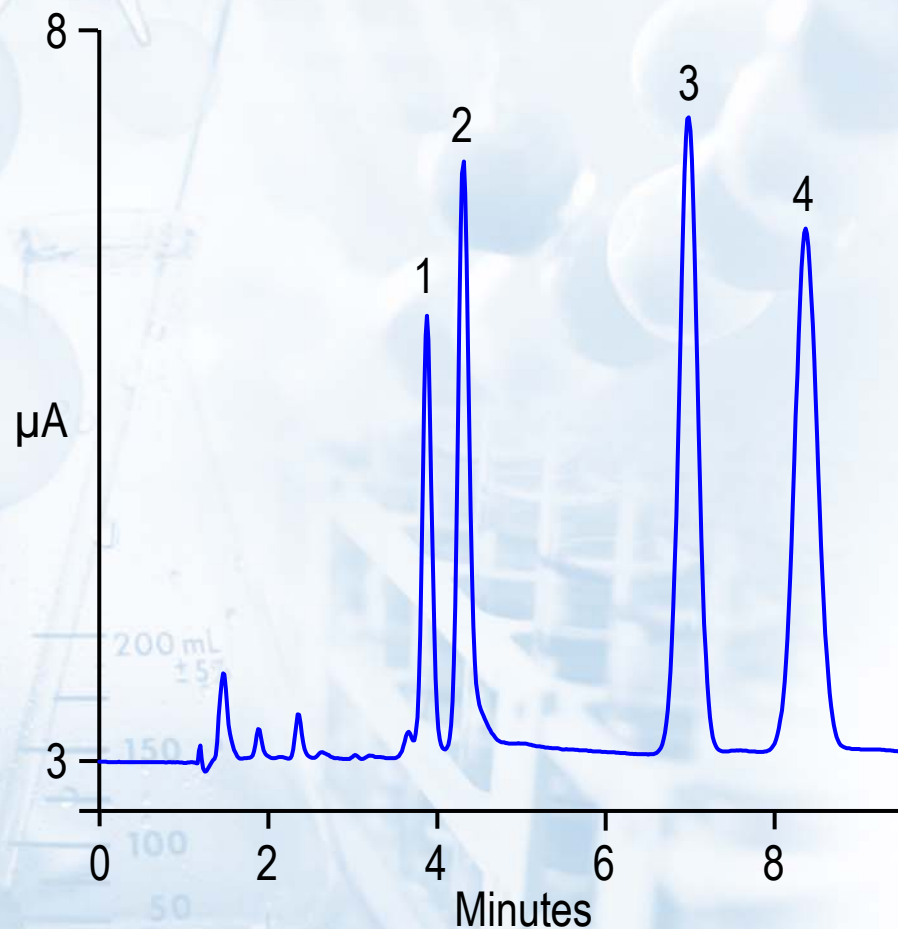
Column: CarboPac™ PA1 and guard

Eluent: 150 mM NaOH

Flow Rate: 1.0 mL/min

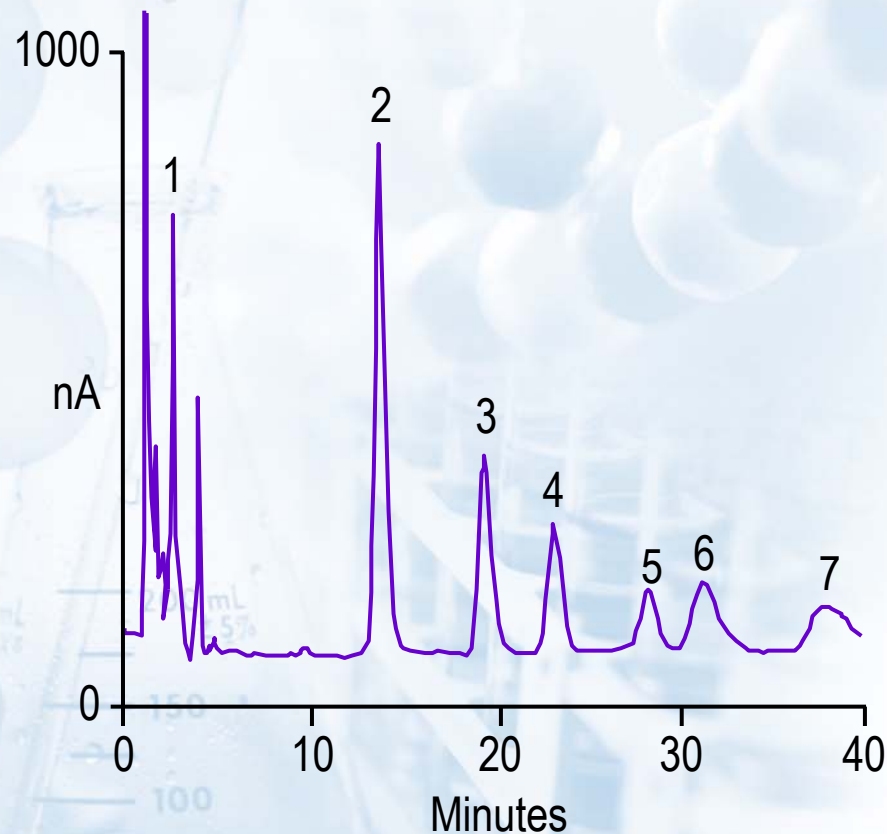
Inj. Volume: 50 μ L

Detection: Pulsed amperometry



Peaks:	1. Glucose	4.39%
	2. Fructose	6.67
	3. Lactose (int. std.)	–
	4. Sucrose	30.8

Carbohydrates in Instant Coffee



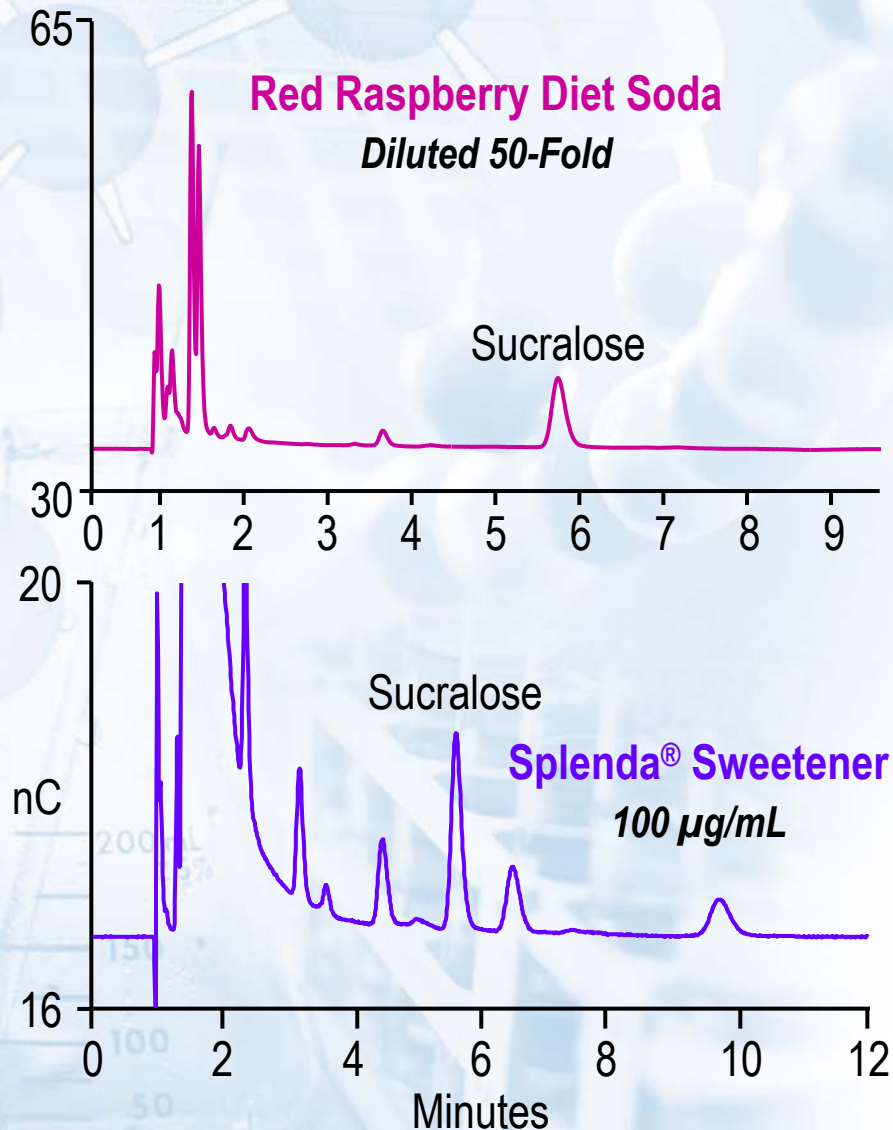
Column: CarboPac™ PA1
Eluent: 150 mM sodium hydroxide/
deionized water gradient
Inj. Volume: 25 μ L of 10 g/L solution
Detector: Pulsed amperometry,
Au electrode; postcolumn
addition of 0.3 M NaOH

Peaks:

1. Mannitol	21 mg/L
2. Arabinose	140
3. Galactose	76
4. Glucose	44
5. Xylose	26
6. Mannose	51
7. Fructose	93

Sample Preparation: Phenolics removed with
OnGuard® P cartridge

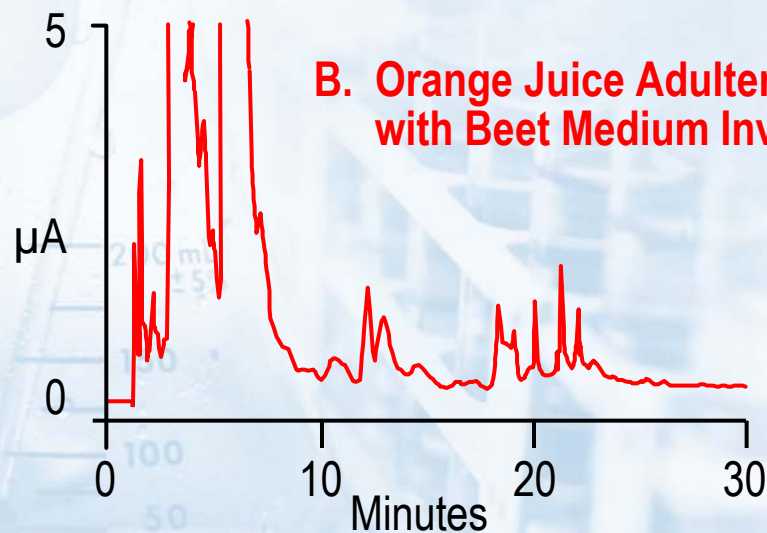
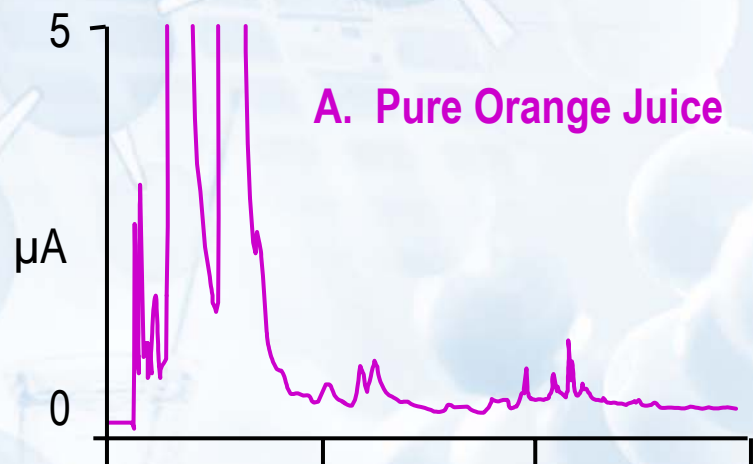
Sucralose in Diet Drink and Artificial Sweetener Product



Column: CarboPac™ PA20 with guard
Eluents: 40 mM sodium hydroxide/
75 mM sodium acetate
Temperature: 30 °C
Flow Rate: 0.5 mL/min, 10-g/L solution
Inj. Volume: 25 µL
Detection: Pulsed amperometric
detection, carbohydrate
waveform, disposable gold
working electrode

Splenda is a registered trademark of McNeil-PP

Oligosaccharide Profiles of Pure and Adulterated Orange Juice

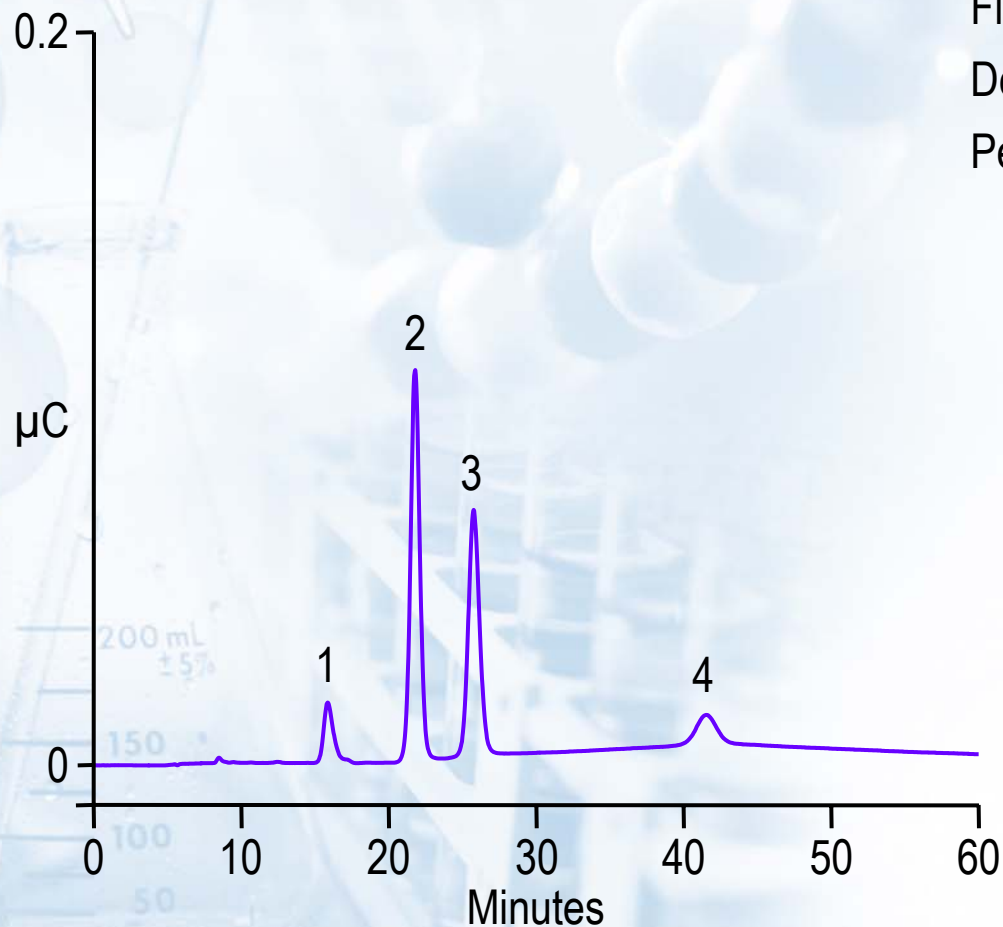


Column: CarboPac™ PA-100
Eluent: Sodium hydroxide/
sodium acetate gradient
Flow Rate: 1 mL/min
Conc.: 2 mg/mL
Inj. Volume: 25 μL
Detection: Pulsed amperometry,
Au electrode

Apple Juice

Column: CarboPac™ MA1
Eluent: 500 mM sodium hydroxide
Flow Rate: 0.4 mL/min
Detection: Pulsed amperometry

Peaks:	1. Sorbitol	14	μg
	2. Glucose	130	
	3. Fructose	36	
	4. Sucrose	0.2	



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